



EXPANDED FUNGAL REPORT®

Prepared Exclusively For

Alpha Check Inspections 581 Lindsey Drive Wayne, PA 19087

Report Date: 5/23/2012

Project: 535 Tree La, West Chester, PA 19380

EMSL Order: 371207865



AIHA-LAP, LLC--EMLAP Lab 100194



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Attn: John Biegalski

Alpha Check Inspections 581 Lindsey Drive Wayne, PA 19087 EMSL Order: 371207865 Customer ID: ALPC34 Collected: 5/21/2012 Received: 5/22/2012 Analyzed: 5/22/2012

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1. Description of Analysis

Analytical Laboratory

EMSL Analytical, Inc. (EMSL) is a nationwide, full service, analytical testing laboratory network providing Asbestos, Mold, Indoor Air Quality, Microbiological, Environmental, Industrial Hygiene and Mechanical Testing services since Chemical, Forensic, Materials, 1981. Ranked as the premier independently owned environmental testing laboratory in the nation, EMSL puts analytical quality as its top priority. This quality is recognized by many well-respected federal, state and private accrediting agencies, such as AIHA's EMLAP and EMPAT programs, and assured by our high quality personnel, including many Ph.D. microbiologists and mycologists.

EMSL is an independent laboratory that performed the analysis of these samples. EMSL did not conduct the sampling or site investigation for this report. The samples referenced herein were analyzed under strict quality control procedures using state-of-the-art microbiological methods. The analytical methods used and the data presented are scientifically and legally defensible.

The laboratory data is provided in compliance with AIHA policy modules and ISO 17025 guidelines for the particular test(s) requested, including any associated limitations for the methods employed. These data are intended for use by professionals having knowledge of the testing methods necessary to interpret them accurately.

Air Samples - Spore traps:

Spore traps are commercially available sampling devices that capture airborne particles on an adhesive slide. Air is pulled through the device using a vacuum pump. Spores, as well as other airborne particles, are impacted on the collection adhesive. Using spore trap collection methods has inherent limitations. These collection methods are biased towards larger spore sizes.

The analysis for total spore counts is a direct microscopic examination and does not include culturing or growing the fungi. Therefore, the results include both viable and non-viable spores. Some fungal groups produce similar spore types that cannot be distinguished by direct microscopic examination alone (i.e., *Aspergillus/Penicillium*, and others). Other spore types may lack distinguishing features that aid in their identification. These types are grouped into larger categories such as Ascospores or Basidiospores.

Fungal spores are identified and grouped by morphological characteristics including color, shape, septation, ornamentation, and fruiting structures (if present) which are compared to published mycological identification keys and texts. EMSL reports provide spore counts per cubic meter of air to three significant figures. Please note that each spore category is reported to three significant figures. Due to rounding and the application of three significant



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figures the sum of the individual spore numbers may not equal the total spore count on the report. EMSL does not maintain responsibility for final volume concentrations (counts/m3) since this volume is provided by the field collector and can not be verified by EMSL.

EMSL analyzes spore traps using phase contrast microscopy. There is a wide choice of collection devices (Air-O-Cell, Micro-5, Burkhard, etc.) on the market. Differences in analytical method may exist between spore trap devices.

Spore trap results are reported in spores per cubic meter of air. Due to the other airborne particles collected with the spores, EMSL reports a background particle density. Background density is an indication of overall particulate matter present on the sample (i.e. dust in the air). High background concentrations may obscure spores such as the *Penicillium/Aspergillus* group. The rating system is from 1-5 with 1 = 1 - 25% of the background obscured by material, 2 = 26 - 50%, 3 = 51 - 75%, 4 = 76% - 99%, 5 = 100% or overloaded. A background rating of 4 or higher should be regarded as a minimum count since the actual concentrations may be higher than those reported. EMSL will not be held responsible for overloading of samples. Sample volumes are left to the discretion of the company or persons conducting the fieldwork.

Skin fragment density is the percentage of skin cells making up the total background material, 1 = 1 - 25%, 2 = 26 - 50%, 3 = 51 - 75%, 4 = 76-100%. Skin fragment density is considered an indication of the general cleanliness in the area sampled. It has been estimated that up to 90% of household dust consists of dead skin cells.

2. Analytical Results

See attached data reports and charts.



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Test Report: Air-O - Cell(™) Analysis of Fungal Spores & Particulates by Optical Microscopy (EMSL Method 05-TP-003)

	Particle Identification	Sample Concentration (counts/m³)	В	ackground Concentration (counts/m³)	Background Corrected (counts/m³)
Lab Camula Number	Alternaria	21		7*	14
Lab Sample Number	Ascospores	63		7960	Less than Background
371207865-0001	Aspergillus/Penicillium	232		None Detected	232
	Basidiospores	190		13200	Less than Background
Client Sample ID	Bipolaris++	None Detected		None Detected	Equal To Background
17443319	Chaetomium	None Detected		21	Less than Background
	Cladosporium	232		1250	Less than Background
	Curvularia	None Detected		None Detected	Equal To Background
Location	Epicoccum	7*		21	Less than Background
	Fusarium	None Detected		None Detected	Equal To Background
2nd Fl Hallway-Indoor	Ganoderma	21	l	None Detected	21
	Myxomycetes++	106		42	64
Sample Volume (L)	Oidium	None Detected	l	7*	Less than Background
450	Pestalotia	13*		None Detected	13
150	Pithomyces	None Detected		None Detected	Equal To Background
	Rust	None Detected		None Detected	Equal To Background
Sample Type	Scopulariopsis	None Detected		None Detected	Equal To Background
Inside	Stachybotrys	None Detected		None Detected	Equal To Background
	Torula	None Detected		None Detected	Equal To Background
Comments	Ulocladium	None Detected		None Detected	Equal To Background
Comments	Unidentifiable Spores	21		None Detected	21
	Zygomycetes	None Detected		None Detected	Equal To Background
	Total Fungi	906		22500	Less than Background
	Hyphal Fragment	106		42	64
	Insect Fragment	21		None Detected	21
	Pollen	63		42	21
	Analytical Sensiti	vity 600x:	21	counts/cubic meter	
	Analytical Sensitivity 300x *:		7*	counts/cubic meter	
	Skin Fragments:		2	1 to 4 (low to high)	
	Fibrous Particulat	te:	1	1 to 4 (low to high)	
	Background:		4	1 to 4 (low to high); 5 (overloaded)

Initial report from: 05/23/2012 10:48:56

Bipolaris++ = Bipolaris/Dreschlera/Exserohilum

Myxomycetes++ = Myxomycetes/Periconia/Smut

No discernable field blank was submitted with this group of samples.

Samples analyzed by EMSL Analytical, Inc. Cinnaminson, NJ AIHA-LAP, LLC--EMLAP Lab 100194

High levels of background particulate can obscure spores and other particulates leading to underestimation. Background levels of 5 indicate an overloading of background particulates, prohibiting accurate detection and quantification. Present = Spores detected on overloaded samples. Results are not blank corrected unless othewise noted. The detection limit is equal to one fungal spore, structure, pollen, fiber particle or insect fragment. "*" Denotes particles found at 300X. "-" Denotes not detected. Due to method stopping rules, raw counts in excess of 100 are extrapolated based on the percentage analyzed. EMSL maintains liability limited to cost of analysis. This report relates only to the samples reported above and may not be reproduced, except in full, without written approval by EMSL. EMSL bears no responsibility for sample collection activities or analytical method limitations. Interpretation and use of test results are the responsibility of the client. Samples received in good condition unless otherwise noted.

Farbod Nekouei, M.S., Laboratory Manager or other Approved Signatory

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		Sample Concentration	Background Concentration	Background Corrected
	Particle Identification	(counts/m³)	(counts/m³)	(counts/m³)
Lab Sample Number	Alternaria	7*	7*	N/A
Lab Cample Humber	Ascospores	7960	7960	N/A
371207865-0002	Aspergillus/Penicillium	None Detected	None Detected	N/A
	Basidiospores	13200	13200	N/A
Client Sample ID	Bipolaris++	None Detected	None Detected	N/A
17443288	Chaetomium	21	21	N/A
	Cladosporium	1250	1250	N/A
	Curvularia	None Detected	None Detected	N/A
Location	Epicoccum	21	21	N/A
	Fusarium	None Detected	None Detected	N/A
Front Porch-Outdoor	Ganoderma	None Detected	None Detected	N/A
	Myxomycetes++	42	42	N/A
Sample Volume (L)	Oidium	7*	7*	N/A
450	Pestalotia	None Detected	None Detected	N/A
150	Pithomyces	None Detected	None Detected	N/A
Sample Type	Rust	None Detected	None Detected	N/A
	Scopulariopsis	None Detected	None Detected	N/A
Background	Stachybotrys	None Detected	None Detected	N/A
	Torula	None Detected	None Detected	N/A
Comments	Ulocladium	None Detected	None Detected	N/A
Comments	Unidentifiable Spores	None Detected	None Detected	N/A
	Zygomycetes	None Detected	None Detected	N/A
	Total Fungi	22500	22500	N/A
	Hyphal Fragment	42	42	N/A
	Insect Fragment	None Detected	None Detected	N/A
	Pollen	42	42	N/A
	Analytical Sensitivity 600x: Analytical Sensitivity 300x *: Skin Fragments:		21 counts/cubic meter	
			7* counts/cubic meter	
			1 1 to 4 (low to high)	
	Fibrous Particulat	e:	1 1 to 4 (low to high)	
	Background:		1 1 to 4 (low to high); 5 (d	overloaded)

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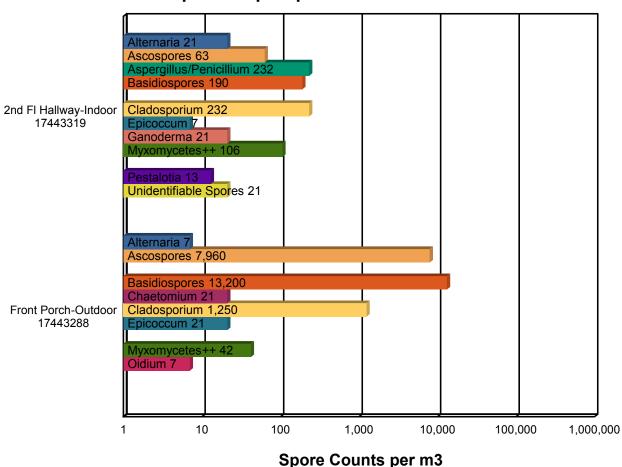
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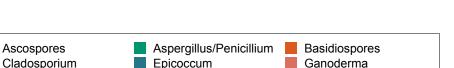
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Spore Trap Report: Total Counts





Unidentifiable Spores

Pestalotia

Oidium

Alternaria

Chaetomium

Myxomycetes++

^{*} The chart is displayed using a logarithmic scale. Bar size is not directly proportional to the number of spores.



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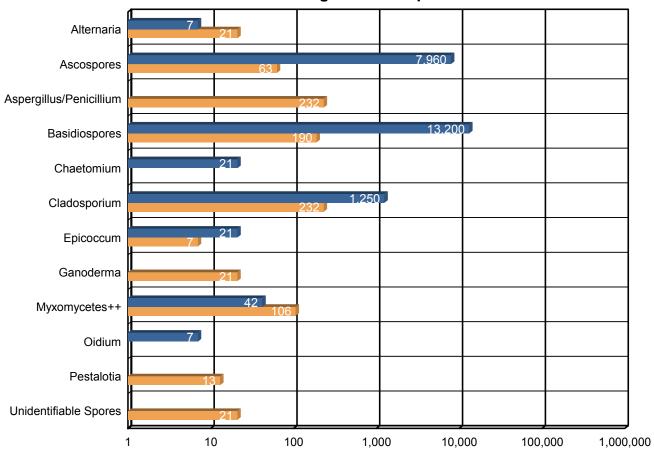
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Background Comparison Chart



Spore Counts per m3



^{*} The chart is displayed using a logarithmic scale. The bar size is not directly proportional to the number of spores.



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Test Report: Microscopic Examination of Fungal Spores, Fungal Structures, Hyphae, and Other Particulates from Tape Samples (EMSL Method: M041)

Lab Sample Number: Client Sample ID: Sample Location:	371207865-0003 53599 Attic Tape Lift				
Spore Types	Category	-	-	-	-
Agrocybe/Coprinus	-				
Alternaria	-				
Ascospores	-				
Aspergillus/Penicillium	Low				
Basidiospores	-				
Bipolaris++	-				
Chaetomium	-				
Cladosporium	*High*				
Curvularia	-				
Epicoccum	-				
Fusarium	-				
Ganoderma	-				
Myxomycetes++	-				
Paecilomyces	-				
Rust	-				
Scopulariopsis	-				
Stachybotrys	-				
Torula	-				
Ulocladium	High				
Unidentifiable Spores	-				
Zygomycetes	-				
Fibrous Particulate	-				
Hyphal Fragment	-				
Insect Fragment	-				
Pollen	-				
Initial report from: 05/23/2012 10:48:56					

Category: Count/per area analyzed

Rare: 1 to 10 Low: 11 to 100 Medium: 101 to 1000 High: >1000

Bipolaris++ = Bipolaris/Dreschlera/Exserohilum Myxomycetes++ = Myxomycetes/Periconia/Smut

* = Sample contains fruiting structures and/or hyphae associated with the spores.

No discernable field blank was submitted with this group of samples.

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3. Understanding the Results

EMSL Analytical, Inc. is an independent laboratory, providing unbiased and scientifically valid results. These data represent only a portion of an overall IAQ investigation. Visual information and environmental conditions measured during the site assessment (humidity, moisture readings, etc.) are crucial to any final interpretation of the results. Many factors impact the final results; therefore, result interpretation should only be conducted by qualified individuals. The American Conference of Governmental Industrial Hygienists (ACGIH) has published a good reference book covering sampling and data interpretation. It is entitled, Bioaerosols: Assessment and Control, 1999.

Air Samples:

Air samples are typically evaluated by means of fungal type identified and by comparing indoor and outdoor concentrations, complaint to non-complaint areas, or area of concern to areas of non-concern. In general, the levels and types of fungi in the indoor air (in non-problem buildings) should be similar to or lower than those found in the outdoor air. Higher levels of spores (order of magnitude) found inside may indicate that moisture sources and resultant fungal growth are present. Spore count results are influenced by geographic location, seasonal and diurnal variation, and biotic/abiotic outside conditions. For example, fresh snow cover on the ground will affect the outdoor spore count concentration.

Use the following guidelines when interpreting the results:

- 1. The composition and diversity of fungi in indoor, non-problem buildings should be similar to that of the outdoor air.
- Compare spore count concentrations indoors and outdoors. Elevated indoor concentrations may indicate indoor fungal growth. Be aware that this is not always consistent and additional sampling may be needed.
- 3. Certain fungi are very good indicators of water damage. The presence of these spores, even in small quantities, indicates the presence of water damage. These indicator fungi include, but are not limited to, *Chaetomium, Fusarium, Stachybotrys* (including *Memnoniella*), and *Ulocladium*.
- 4. Different types of fungi grow at different levels of biologically available water. These differences in fungal growth suggest the degree of water damage or saturation. For example, *Stachybotrys* is an indication of short term, severe, or prolonged water damage over time.



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Fungal spores are found everywhere. Whether or not symptoms develop in people exposed to fungi depends on the nature of the fungal material (e.g., allergenic, toxic, or infectious), the exposure level, and the susceptibility of exposed persons. Susceptibility varies with the genetic predisposition (e.g., allergic reactions do not always occur in all individuals), age, pre-existing medical conditions (e.g., diabetes, cancer, or chronic lung conditions), use of immunosuppressive drugs, and concurrent exposures. These reasons make it difficult to identify dose/response relationships that are required to establish "safe" or "unsafe" levels (i.e., permissible exposure limits).

It is generally accepted in the industry that indoor fungal growth is undesirable and inappropriate, necessitating removal or other appropriate remedial actions. The New York City guidelines and EPA guidelines for mold remediation in schools and commercial buildings define the conditions warranting mold remediation. Always remember that water is the key. Preventing water damage or water condensation will prevent mold growth.

This report is not intended to provide medical advice or advice concerning the relative safety of an occupied space. Always consult an occupational or environmental health physician who has experience addressing indoor air contaminants if you have any questions.



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4. Glossary of Fungi

ALTERNARIA	
Natural Habitat	Common saprobe and pathogen of plants. Typically found on plant tissue, decaying wood, and
	foods. Soil . Air outdoors.
Suitable Substrates in the	Indoors near condensation (window frames, showers), House dust (in carpets, and air). Also
Indoor Environment	colonizes building supplies, computer disks, cosmetics, leather, optical instruments, paper,
	sewage, stone monuments, textiles, wood pulp, and jet fuel
Water Activity	Aw =0.85-0.88
Mode of Dissemination	Wind
Allergic Potential	Type I allergies (hay fever, asthma), Type III (hypersensitivity pneumonitis)
Potential or Opportunistic	Phaeohyphomycosis {causing cystic granulomas in the skin and subcutaneous tissue}. In
Pathogens	immunocompetent patients, Alternaria colonizes the paranasal sinuses, leading to chronic
	hypertrophic sinusitis
Industrial Uses	Biocontrol of weed plants ·Biocontrol fungal plant pathogens.
Potential Toxins Produced	Alternariol (AOH) . Alternariol monomethylether (AME). Tenuazonic acid (TeA). Altenuene
	(ALT). Altertoxins (ATX)
Other Comments	Alternaria spores are one of the most common and potent indoor and outdoor airborne
	allergens. Additionally, Alternaria sensitization has been determined to be one of the most
	important factors in the onset of childhood asthma. Synergy with Cladosporium or Ulocladium
	may increase the severity of symptoms
	I may indease the seventy of symptoms



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ASCOSPORES	
Natural Habitat	Everywhere in nature.
Suitable Substrates in the	Depends on genus and species.
Indoor Environment	
Water Activity	Depends on genus and species.
Mode of Dissemination	Forcible ejection or passive release and dissemination by wind or insects.
Allergic Potential	Depends on genus and species.
Potential or Opportunistic	Depends on genus and species.
Pathogens	
Industrial Uses	Depends on genus and species.
Potential Toxins Produced	Depends on genus and species.
Other Comments	Ascospores are the result of sexual reproduction and produced in a saclike structure called an ascus. All ascospores belong to members of the Phylum Ascomycota, which encompasses a plethora of genera worldwide.



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ASPERGILLUS/PENIC	ASPERGILLUS/PENICILLIUM		
Natural Habitat	·Plant debris ·Seed ·Cereal crops		
Suitable Substrates in the Indoor Environment	Grows on a wide range of substrates indoors ·Prevalent in water damaged buildings ·Foods (blue mold on cereals, fruits, vegetables, dried foods) ·House dust ·Fabrics ·Leather ·Wallpaper ·Wallpaper glue		
Water Activity	Aw=0.75-0.94		
Mode of Dissemination	Wind Insects		
Allergic Potential	Type I (hay fever, asthma) ·Type III (hypersensitivity)		
Potential or Opportunistic Pathogens	Possible depending on the species.		
Industrial Uses	Many depending on the species		
Potential Toxins Produced	Possible depending on the species.		
Other Comments	Spores of Aspergillus and Penicillium (including others such as Acremonium and Paecilomyces) are small and spherical with few distinguishing characteristics. They cannot be differentiated or speciated by non-viable impaction sampling methods. Some species with very small spores may be undercounted in samples with high background debris.		



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BASIDIOSPORES		
Natural Habitat	Forest floors. Lawns .Plants (saprobes or pathogens depending on genus)	
Suitable Substrates in the Indoor Environment	Depends on genus. Wood products	
Water Activity	Unknown.	
Mode of Dissemination	Forcible ejection. Wind currents.	
Allergic Potential	Type I allergies (hay fever, asthma) . Type III (hypersensitivity pneumonitis)	
Potential or Opportunistic Pathogens	Depends on genus.	
Industrial Uses	Edible mushrooms are used in the food industry.	
Potential Toxins Produced	Amanitins. monomethyl-hydrazine. muscarine. ibotenic acid. psilocybin.	
Other Comments	Basidiospores are the result of sexual reproduction and formed on a structure called the basidium. Basidiospores belong to the members of the Phylum Basidiomycota, which includes mushrooms, shelf fungi, rusts, and smuts.	



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CHAETOMIUM	
Natural Habitat	Dung. Seeds. Soil. Straw.
Suitable Substrates in the Indoor Environment	Paper. Sheetrock. Wallpaper.
Water Activity	Aw=0.84-0.89.
Mode of Dissemination	Wind. Insects. Water splash.
Allergic Potential	Type I (asthma and hay fever).
Potential or Opportunistic Pathogens	Onychomycosis. C. perlucidum recognized as a new agent of cerebral phaeohyphomycosis.
Industrial Uses	Cellulase production, Textile testing.
Potential Toxins Produced	Chaetomin. Chaetoglobosins A,B,D and F are produced by Chaetomium globosum. Sterigmatocystin is produced by rare species



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CLADOSPORIUM	
Natural Habitat	Dead plant matter. Straw. Soil. Woody plants
Suitable Substrates in the Indoor Environment	Fiberglass duct liner. Paint. Textiles. Found in high concentration in water-damaged building materials.
Water Activity	Aw 0.84-0.88
Mode of Dissemination	Air
Allergic Potential	Type I (asthma and hay fever).
Potential or Opportunistic	Edema. keratitis. onychomycosis. pulmonary infections. Sinusitis.
Pathogens	
Industrial Uses	Produces 10 antigens.
Potential Toxins Produced	Cladosporin and Emodin.



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EPICOCCUM		
Natural Habitat	A worldwide saprophytic fungi, being isolated from dead plant material and soil.	
Suitable Substrates in the	Paper, textiles	
Indoor Environment		
Water Activity	0.86-0.90	
Mode of Dissemination	Wind	
Allergic Potential	Hay fever, asthma	
Potential or Opportunistic	Unknown	
Pathogens		



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GANODERMA	
Natural Habitat	Grows on conifers and hardwoods worldwide, causing white rot, root rot, and stem rot.
Suitable Substrates in the	Unknown.
Indoor Environment	
Water Activity	Unknown.
Mode of Dissemination	Wind.
Allergic Potential	Ganoderma species are known to cause allergies in people on a worldwide scale.
Potential or Opportunistic	Unknown.
Pathogens	
Industrial Uses	Biopulping of wood for the paper industry. Potential medicinal use due to: 1. Inhibition of Ras
	dependent cell transformation, 2. Antifibrotic activity, 3. Immunomodulating activity, 4.
	Free-radicle scavenging
Potential Toxins Produced	Unknown.
Other Comments	Used in traditional Chinese medicine as an herbal supplement. It is also known as a "shelf
	fungus" because the fruiting body forms a stalk-less shelf on the sides of trees and logs. It is
	sometimes called "artists conk" because when you scratch the white pores of the fruiting body,
	the white rubs away and exposes the brown hyphae underneath. Thus, pictures can be
	produced on the fruiting body.
Reference	References: Craig, R.L., Levetin, E. 2000. Multi-year study of Ganoderma aerobiology.
	Aerobiologia 16: 75-81.
	http://www.pfc.forestry.ca/diseases/CTD/Group/Heart/heart6_e.html



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MYXOMYCETES++	
Natural Habitat	Decaying logs, Dead leaves , Dung , Lawns , Mulched flower beds, Lawns
Suitable Substrates in the Indoor Environment	Rotting lumber
Free moisture required for mold growth	Unknown
Mode of Dissemination	Insects, Water, Wind
Allergic Potential	Type I
Potential or Opportunistic	Unknown
Pathogens	
Industrial Uses	



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ULOCLADIUM	
Natural Habitat	A worldwide saprophytic fungi, being isolated from dead plant material and soil.
Suitable Substrates in the	Wallboard, jute, paper, wood, textiles. Good indicator of water damage.
Indoor Environment	
Water Activity	0.89
Mode of Dissemination	Wind
Allergic Potential	Hay fever, asthma
Potential or Opportunistic	Unknown
Pathogens	



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5. References and Informational Links

Books

- Bioaerosols: Assessment and Control. Janet Macher, Ed., American Conference of Governmental Industrial Hygienists, Cincinnati, OH 1999.
- Exposure Guidelines for Residential Indoor Air Quality. Environmental Health Directorate, Health Protection Branch, Health Canada, Ottawa, Ontario, 1989.
- Fungal Contamination in Public Buildings: Health Effects and Investigation Methods.
 Health Canada, Ottawa, Ontario, 2004.
- IICRC: S500 Standard and Reference Guide for Professional Water Damage Restoration. 3rd Edition, Institute of Inspection, Cleaning, and Restoration Certification, Vancouver, WA, 2006
- IICRC: S520 Standard and Reference Guide for Professional Mold Remediation. 1st Edition, Institute of Inspection, Cleaning, and Restoration Certification, Vancouver, WA, 2004
- Field Guide for the Determination of Biological Contaminants in Environmental Samples.
 2nd Edition, American Industrial Hygiene Association, 2005.

Consumer Links

- Read the full text of AIHA's "The Facts About Mold" consumer brochure.
 http://www.aiha.org/news-pubs/newsroom/Documents/Facts%20about%20mold.pdf
- The Occupational Safety and Health Administration (OSHA) http://www.osha.gov/SLTC/molds/index.html
- CDC Mold Facts <u>http://www.cdc.gov/mold/faqs.htm</u>
- CDC Stachybotrys Questions and answers on Stachybotrys chartarum and other molds http://www.cdc.gov/mold/stachy.htm
- IOM, NAS: Clearing the Air: Asthma and Indoor Air Exposures http://fermat.nap.edu/books/0309064961/html/index.html
- National Library of Medicine-Mold website http://www.nlm.nih.gov/medlineplus/molds.html
- California Department of Health Services (CADOHS)



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http://www.cal-iag.org/mold/about-mold

Minnesota Department of Health http://www.health.state.mn.us/divs/eh/indoorair/mold/index.html

- New York City Department of Health and Mental Hygiene http://www.nyc.gov/html/doh/html/epi/moldrpt1.shtml
- H.R.: The United States Toxic Mold Safety and Protection Act http://conyers.house.gov/index.cfm?FuseAction=Issues.Home&Issue id=061bf20d-19b9 -b4b1-12a3-6089055e7c99

EPA

"Should You Have the Air Ducts in Your Home Cleaned?"

http://www.epa.gov/iaq/pubs/flood.html

General information about molds and actions that can be taken to clean up or prevent a mold problem.

http://www.epa.gov/asthma/molds.html

- "A Brief Guide to Mold, Moisture, and Your Home" Includes basic information on mold, cleanup guidelines, and moisture and mold prevention. http://www.epa.gov/iag/molds/moldguide.html
- "Mold Remediation in Schools and Commercial Buildings" Information on remediation in schools and commercial property, references for potential mold and moisture remediators.

http://www.epa.gov/mold/mold_remediation.html

FEMA

"Homes That Were Flooded May Harbor Mold Problems" - Information and tips for cleaning mold.

http://www.fema.gov/diz01/d1364n18.shtm

- "Dealing With Mold & Mildew in Your Flood Damaged Home. http://www.fema.gov/pdf/rebuild/recover/fema mold brochure english.pdf
- "Prompt Flood Cleanup Can Help Prevent Health Problems" How to clean up in-house



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mold problems (not large or serious exposures).

http://www.fema.gov/news/newsrelease.fema?id=9538



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Samples analyzed by EMSL will be retained for 60 days after analysis date Storage beyond this period is available for a fee with written request prior to the initial 30 day period. Samples containing hazardous/toxic substances which require special handling will be returned to the client immediately. EMSLreserves the right to charge a sample disposal fee or return samples to the client.

B. Change Orders and Cancellation

All changes in the scope of work or turnaround time requested by the client after sample acceptance must be made in writing and confirmed in writing by EMSL. If requested changes result in a change in cost the client must accept payment responsibility. In the event work is cancelled by a client, EMSL will complete work in progress and invoice for work completed to the point of cancellation notice. EMSL is not responsible for. holding times that are exceeded due to such changes.

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